



## Relevant Gene Content

The content for TruSight Tumor 15 focuses on relevant regions in 15 cancer-associated genes (Table 1). The gene content was carefully selected to include content cited by industry organizations such as the National Comprehensive Cancer Network (NCCN)<sup>3</sup> and the European Society for Medical Oncology (ESMO).<sup>4</sup> Independent consortia publications and late-stage pharmaceutical research also influenced the design of TruSight Tumor 15.<sup>5-7</sup> These genes and gene regions include single nucleotide variants (SNV) and insertions and deletions (indels) that have demonstrated involvement in solid tumors. By harnessing the expertise of recognized authorities in the oncology community, TruSight Tumor 15 enables researchers to focus resources on relevant genes that are most likely to play a role in tumorigenesis.

**Table 1: Gene Content on TruSight Tumor 15**

<i>AKT1</i>	<i>GNA11</i>	<i>NRAS</i>
<i>BRAF</i>	<i>GNAQ</i>	<i>PDGFRA</i>
<i>EGFR</i>	<i>KIT</i>	<i>PIK3CA</i>
<i>ERBB2</i>	<i>KRAS</i>	<i>RET</i>
<i>FOXL2</i>	<i>MET</i>	<i>TP53</i>

## Optimized for Low DNA Input and FFPE Tissue

Archival FFPE tumor samples often contain degraded DNA that introduces data inaccuracies, and the extraction process yields small amounts of usable DNA for NGS. The library preparation and sequencing methods for TruSight Tumor 15 are designed to address these challenges. Sample quantification guidelines are provided to ensure reliable, high-quality sequencing data from FFPE tissue. By maximizing sample success rates across multiple tumor types, the panel enables conservation of limited samples and resources.

**Table 2: Specifications**

Parameter	Details
Panel Size	44 kb
Content	250 amplicons
Amplicon Size	Average ~150–175 bp
DNA Input Requirement	20 ng total (10 ng × 2 reactions)
Library Preparation Time	7 hours total time, 3.5 hours hands-on time
Sequence Run Time	24 hours (MiniSeq) or 27 hours (MiSeq)
Sequence Run	2 × 151 bp
Sample Throughput	8 samples per run using MiniSeq High Throughput Kit
	8 samples per run using MiSeq v3 chemistry
Variation Frequency	5%
Amplicon Coverage	93.5% of bases covered at ≥ 500×

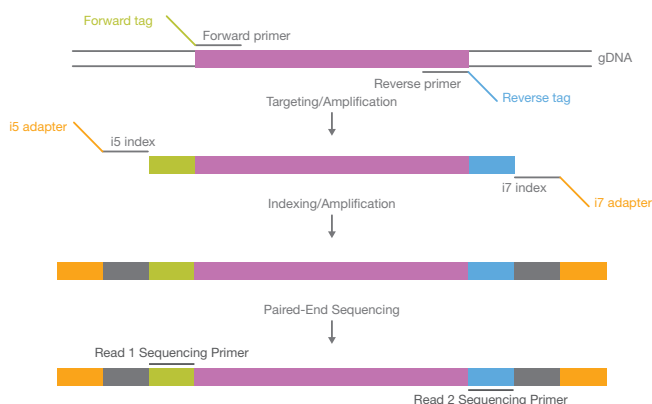
## Sensitive, High-Confidence Variant Detection

Deep sequencing using NGS provides the high sensitivity to reveal somatic variation in tumor subpopulations. Illumina sequencing by synthesis (SBS) chemistry is the most widely adopted NGS technology, generating > 90% of global sequencing data.\* When paired with high-quality sequencing on the MiSeq Systems,<sup>7,8</sup> TruSight Tumor 15 provides uniform coverage of target regions, identifying somatic

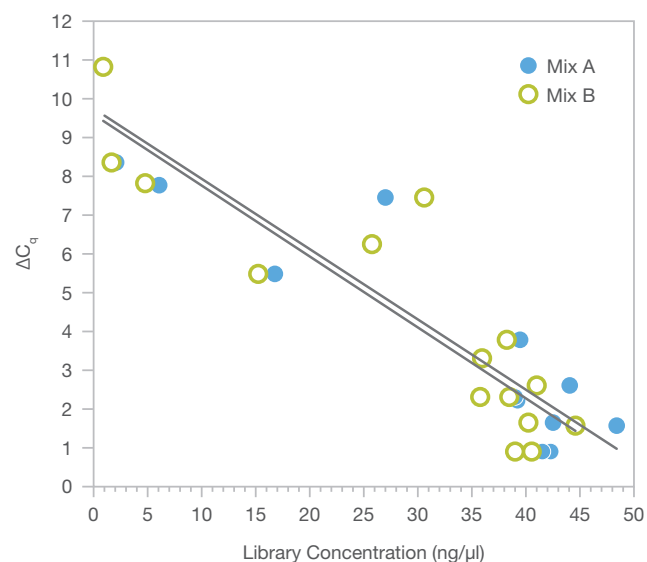
\* Data calculations on file. Illumina, Inc., 2015

mutations as low as 5% variant allele frequency with ≥ 500× minimum coverage (Table 2).

The TruSight Tumor 15 library preparation assay follows a tiling method that uses 2 oligo pools for multiplex PCR (Figure 2). This strategy enables coverage of larger DNA regions, produces higher coverage uniformity, and reduces the presence of primer dimers and FFPE-induced artifacts. This results in high accuracy and sensitivity. Instead of requiring a separate, preliminary qPCR reaction, inprocess quality recommendations are provided during library preparation process, supporting sample success (Figure 3). These quality recommendations include final library concentration and expected library size.



**Figure 2: TruSight Tumor 15 Chemistry**—The TruSight Tumor 15 assay uses a multiplex PCR approach, resulting in high accuracy and sensitivity.



**Figure 3: Concordance of Quality Metrics**— The quality threshold used for TruSight Tumor 15 (20 ng/µl) is highly correlated with  $\Delta C_q$  results from DNA quality evaluation using qPCR. Mix A and Mix B denote the 2 different libraries prepared per sample.

**Table 3: TruSight Tumor 15 Coverage**

Sample ID	Sample Quality	% of Bases $\geq 500\times$	Amplicon Mean Coverage
FFPE_Colon1	Medium	99.7%	24,219x
FFPE_Colon2	Low	99.9%	20,763x
FFPE_Colon3	Low	99.2%	35,270x
FFPE_Colon4	High	100.0%	18,357x
FFPE_Colon5	High	100.0%	15,769x
FFPE_Melanoma1	Medium	99.7%	32,707x
FFPE_Melanoma2	Low	99.1%	41,640x
FFPE_Melanoma3	High	100.0%	17,285x
FFPE_Melanoma4	Low	95.7%	10,177x
FFPE_Breast1	High	99.1%	15,501x

DNA was extracted from FFPE tumor samples and then 20 ng of input DNA was evaluated using the TruSight Tumor 15 assay and sequenced on the MiniSeq System. Coverage of  $\geq 500\times$  is required for accurate identification of mutations at 5% variant frequency. Sample quality was determined by amplification potential of extracted DNA compared to a non-FFPE control sample in a qPCR assay. High quality is indicated by  $\Delta Cq$  of 0–2. Medium quality is indicated by  $\Delta Cq$  value of 2–4. Low quality is indicated by  $\Delta Cq$  of 4–6.

**Table 4: TruSight Tumor 15 Performance with Characterized Horizon Sample**

Gene	Mutation	Reported Frequency	Detected Frequency	Coverage
<i>BRAF</i>	V600E	10.5%	12.3%	55,457x
<i>KIT</i>	D816V	10.0%	10.3%	5463x
<i>EGFR</i>	$\Delta E746-A750$	2.0%	2.1%	3553x
<i>EGFR</i>	L858R	3.0%	4.1%	1761x
<i>EGFR</i>	T790M	1.0%	1.2%	18,927x
<i>EGFR</i>	G719S	24.5%	25.6%	41,805x
<i>KRAS</i>	G13D	15.0%	15.3%	6745x
<i>KRAS</i>	G12D	6.0%	7.2%	6742x
<i>NRAS</i>	Q61K	12.5%	11.2%	13,154x
<i>PIK3CA</i>	H1047R	17.5%	18.8%	21,522x
<i>PIK3CA</i>	E545K	9.0%	7.8%	13,250x

DNA from the HD-C749 formalin-fixed cell line (Horizon Diagnostics) containing known variants was evaluated using the TruSight Tumor 15 assay and sequenced on the MiniSeq System. Variants were analyzed using VariantStudio. HD-C749 showed 100% concordance over 7 different runs.

**Table 5: TruSight Tumor 15 Performance with FFPE Tumor Samples**

Sample	Reported Mutation	Detected Mutation	Detected Frequency	Coverage
FFPE_Colon1	<i>KRAS</i> G12S	<i>KRAS</i> G12S	22.3%	21,134x
FFPE_Colon2	<i>KRAS</i> G12D	<i>KRAS</i> G12D	11.5%	4322x
FFPE_Colon3	<i>BRAF</i> V600E	<i>BRAF</i> V600E	25.5%	140,040x
FFPE_Colon4	<i>KRAS</i> G12V	<i>KRAS</i> G12V	33.4%	5256x
FFPE_Colon5	<i>KRAS</i> G13D	<i>KRAS</i> G13D	33.0%	4156x
FFPE_Melanoma1	<i>BRAF</i> V600E	<i>BRAF</i> V600E	65.7%	106,924x
FFPE_Melanoma2	<i>KRAS</i> G12R	<i>KRAS</i> G12R	4.1%	54,622x
FFPE_Melanoma3	<i>BRAF</i> V600E	<i>BRAF</i> V600E	93.5%	61,838x
FFPE_Melanoma4	<i>BRAF</i> V600K	<i>BRAF</i> V600K	22.2%	8075x
FFPE_Breast1	<i>AKT1</i> E17K	<i>AKT1</i> E17K	37.3%	56,438x

DNA from FFPE tumor samples was extracted and then evaluated using the TruSight Tumor 15 assay and sequenced on the MiniSeq System. Variants were analyzed using VariantStudio. All 10 FFPE samples had 100% variant concordance.

AAAGAATGATAACAGTAAACACACTTCTGTAACTTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAATTGAGACTAAATATTAACGTACCATTAAAGACTACCGTCTTCTGTAACTTAAAGATTACTTGATCCACTGATTCA/ AATCAACGTACCGTAACGAACGATATATTAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTGAACGAACGAAAAGATTGATAACAGTAAACACACTTCTGTAAAC/ AACGACGAAAAGAATGATAACAGTAAACACACTTCTGTAACTTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTGAACGAACGAAAAGAATGATAAC/ TTTAAGGTACCATTAAAGACTACCGTGAACGAACACTTCTGTAACTTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTGAACGAACGAAAAGAATGATAAC/ AAAGAATGATAACAGTAAACACACTTCTGTAACTTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTCTTCTGTAACTTAAAGATTACTTGATCCACTGATTCA/ AAGATTACTTGATCCACTGATTCAACGTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTGAACGAACGAAAAGAATGATAAC/ AACGTATCAATTGAGACTAAATATTAACGTACCATTAAAGACTCTGTAACTTAAAGATTACTTGATCCACTGATTCAACGTACCGTAACGAACGATCAAITGAGACTAAATATTAACGTACCATTAAAGACTACCGTGAACGAACGAAAAGAATGATAAC/

