Infinium XT - ST Workflow Checklist (no Illumina LIMS)

Amplify DNA (Pre-Amp)

- ☐ 1 Select **MSA7 HT Tasks | Make MSA7 HT**.
  - ☐ a Select the DNA plate type.
- ☐ 2 Place MA1, MA2, and RAM into the tube rack and remove the caps.
- ☐ 3 Add 0.1 N NaOH to a quarter reservoir (5 ml per plate).
- ☐ 4 Place the DNA plates and MSA7 plates on the robot deck.
- ☐ 5 Select **Run**.
- ☐ 6 Vortex the MSA7 plates at 1600 rpm for 1 minute.
- ☐ 7 Centrifuge at 280 × g at room temperature for 1 minute.

Incubate DNA

- ☐ 1 Incubate the MSA7 plates for 3–24 hours at 37°C.

Fragment DNA

- ☐ 1 Centrifuge the MSA7 plates at 280 × g at room temperature for 1 minute.
- ☐ 2 Select **MSA7 HT Tasks | Fragment MSA7 HT**.
- ☐ 3 Place the MSA7 plates on the robot deck.
- ☐ 4 Place FMS tubes into the tube rack and remove the caps.
- ☐ 5 Select **Run**.
- ☐ 6 Select **OK**.
- ☐ 7 Vortex at 1600 rpm for 1 minute.
- ☐ 8 Centrifuge at 280 × g at room temperature for 1 minute.
- ☐ 9 Incubate at 37°C for 30 minutes.

Precipitate DNA

- ☐ 1 Select **MSA7 HT Tasks | Precip MSA7 HT**.
- ☐ 2 Place the MSA7 plates on the robot deck.
- ☐ 3 Add PM1 to a quarter reservoir:

Reagent	Number of Plates	Volume
PM1	1	8 ml
	2	14 ml
	3	21 ml
	4	27 ml
	5	34 ml
	6	40 ml

- ☐ 4 Add 2-propanol to a full reservoir:

Reagent	Number of Plates	Volume
2-propanol	1	25 ml
	2	50 ml
	3	75 ml
	4	100 ml
	5	125 ml
	6	150 ml

- ☐ 5 Select **Run**.
  - ☐ a Select **OK**.
- ☐ 6 Invert the plates 10 times.
- ☐ 7 Centrifuge at 3000 × g at 4°C for 20 minutes.
- ☐ 8 Invert the plates, and drain the supernatant.
- ☐ 9 Tap the plates several times.
- ☐ 10 Air dry for 15 minutes.

Resuspend DNA

- ☐ 1 Select **MSA7 HT Tasks | Resuspend MSA7 HT**.
- ☐ 2 Place the MSA7 plates on the robot deck.
- ☐ 3 Add RA1 to a quarter reservoir.

Reagent	Number of Plates	Volume
RA1	1	5 ml
	2	8 ml
	3	11 ml
	4	14 ml
	5	17 ml
	6	20 ml

- ☐ 4 Apply foil heat seals to the MSA7 plates.
- ☐ 5 Incubate for 15 minutes at 48°C.
- ☐ 6 Vortex at 1800 rpm for 1 minute.
- ☐ 7 Centrifuge at 280 × g at room temperature for 1 minute.

SAFE STOPPING POINT

If you are stopping, store sealed MSA7 plate(s) at 2°C to 8°C for up to 24 hours. If more than 24 hours, store at -25°C to -15°C.

Store sealed RA1 at -25°C to -15°C. If RA1 will be used the next day, seal it, and store it overnight at 4°C.

Hybridize to BeadChip

- ☐ 1 Incubate the MSA7 plates at 95°C for 20 minutes.
- ☐ 2 Cool at room temperature for 30 minutes.
- ☐ 3 Centrifuge at 15001000 × g at room temperature for 1 minute.
- ☐ 4 Place the gaskets into the XT Hyb chambers.
- ☐ 5 Dispense 800 µl PB2 into each reservoir.
- ☐ 6 Close the XT Hyb chamber.
- ☐ 7 Remove all BeadChips from packaging.
- ☐ 8 Place up to 2 BeadChips onto each XT dual Hyb insert and baseplate.
- ☐ 9 Select **MSA7 HT Tasks | Hyb Multi-BC2**.
- ☐ 10 Select the 96-sample BeadChip.
- ☐ 11 Enter the **Number of MSA7 plates**.
- ☐ 12 Place the XT dual Hyb insert and baseplates onto the robot deck.
- ☐ 13 Place the MSA7 plates onto the robot deck.
- ☐ 14 Place an XT tip guide #1 on top of each XT dual Hyb insert and baseplate.
- ☐ 15 Click **Run**, then click **OK**.
- ☐ 16 Remove XT tip guide #1 and replace it with XT tip guide #2, then click **OK**.
- ☐ 17 Remove XT tip guide #2 and replace it with XT tip guide #3, then click **OK**.
- ☐ 18 Click **OK**.
- ☐ 19 Remove XT tip guide #3.
- ☐ 20 Inspect the BeadChips.
- ☐ 21 Load the XT dual Hyb insert and baseplates inside the XT Hyb chambers.
- ☐ 22 Incubate at 48°C for 16 to 24 hours.

Prepare for Next Day

- ☐ 1 Add 330 ml 100% EtOH to the XC4 bottle and shake.
- ☐ 2 Leave the bottle upright on the lab bench overnight.
- ☐ 3 Soak the EXXT tip guides in 1% aqueous Alconox solution.
- ☐ 4 Rinse and dry the EXXT tip guides.

Wash BeadChips

- ☐ 1 Submerge the wash rack in the 1X PB1 wash.
- ☐ 2 Remove the hybridization insert and baseplates.
- ☐ 3 Remove the BeadChips.
- ☐ 4 Remove the cover seals from the BeadChips.
- ☐ 5 Place the BeadChips into the submerged wash rack.
- ☐ 6 Move the wash rack up and down for 1 minute.
- ☐ 7 Move the wash rack to the next 1X PB1 Wash.
- ☐ 8 Move the wash rack up and down for 1 minute.
- ☐ 9 Fill the XCG Flow-Through Chamber assembly tray with 1X PB1.
- ☐ 10 Place a BeadChip on a submerged XCG Flow-Through Chamber frame.
- ☐ 11 Place an XCG glass back plate onto a submerged BeadChip.
- ☐ 12 Attach XCG Flow-Through Chamber clips to each XCG Flow-Through Chamber frame.

Extend and Stain (XStain)

- ☐ 1 Fill the water circulator.
- ☐ 2 Select **Robot QC Tasks | Circulator Manager** to set to 44°C.
- ☐ 3 Select **XStain Tasks | XStain XCG BeadChip ST**.
- ☐ 4 Turn on the iScan systems.
- ☐ 5 Add the following reagents to reservoirs:

Reagent	# BeadChips	Volume
95% formamide/1 mM EDTA	1–8	15 ml
	9–16	17 ml
RA1	1–8	10 ml
	9–16	20 ml
XC3	1–8	50 ml
	9–16	100 ml

- ☐ 6 Invert the LX1, LX2, EML, SML, and ATM tubes to mix. Remove the caps, and place on the robot deck.
- ☐ 7 Enter the number of BeadChips.
- ☐ 8 Select **Run**.
- ☐ 9 Enter the stain temperature listed on the XStain plate.
- ☐ 10 Place the XCG Flow-Through Chamber assemblies into the chamber rack.
- ☐ 11 Select **OK**.
- ☐ 12 Remove the XCG Flow-Through Chamber assemblies from the chamber rack.
- ☐ 13 Set up PB1 and XC4 wash dishes.
- ☐ 14 Pour 310 ml PB1 into a wash dish.
- ☐ 15 Disassemble each XCG flow-through chamber.
- ☐ 16 Place BeadChips into a staining rack in the PB1 wash dish.

- ☐ 17 Submerge the XCG glass back plates in the DI H_2O wash basin.
- ☐ 18 Move the staining rack up and down 10 times.
- ☐ 19 Soak the BeadChips for 5 minutes.
- ☐ 20 Shake the XC4 bottle vigorously.
- ☐ 21 Pour 310 ml XC4 into a wash dish.
- ☐ 22 Move the staining rack to the XC4 wash dish.
- ☐ 23 Move the staining rack up and down 10 times.
- ☐ 24 Soak the BeadChips for 5 minutes.
- ☐ 25 Remove the staining rack.
- ☐ 26 Dry the BeadChips for 50–55 minutes at 675 mm Hg (0.9 bar).
- ☐ 27 Image the BeadChips immediately, or store them, protected from light.